

Phylogenetic Relationships among Bufonoid Frogs (Anura: Neobatrachia) Inferred from Mitochondrial DNA Sequences

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Nucleotide sequences of portions of the mitochondrial 12S and 16S ribosomal RNA genes were used to extend a recent study of anuran phylogeny (Hay *et al.*, *Mol. Biol. Evol.* 12: 928–937, 1995) and to further evaluate phylogenetic relationships within the Neobatrachia. An analysis of almost 900 nucleotides from each of 8 new representatives of the Dendrobatidae, Hylidae, Leptodactylidae, and Myobatrachidae, plus 14 available members of the Neobatrachia provides support for 2 major lineages (Bufonoidea and Ranoidea) within this anuran suborder. The neotropical Bufonoidea and their derivatives are monophyletic. There is an interesting association of the 2 Australian myobatrachids with the South African Heleophryniidae, and the Sooglossidae is one of the basal bufonoid lineages. Within the New World bufonoid frogs, a monophyletic Dendrobatidae is strongly supported. An Australian hylid (Pelodyadinae) shows close affinity with the South American hylid Phyllomedusinae. A group composed of Hylinae (*Hyla* and *Smilisca*), Centrolenidae, Bufonidae, and the hylid Hemiphractinae, with the latter two clustered, was supported significantly. The addition of new taxa has more clearly defined some relationships within the suborder Neobatrachia and has indicated that the families Hylidae, Leptodactylidae, and Myobatrachidae may not be monophyletic. © 1996 Academic Press, Inc.

INTRODUCTION

Among living amphibians the vast majority (88%) of the more than 4500 species belong to the order Anura,

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most of which (96%) are classified as advanced frogs (suborder: Neobatrachia). Phylogenetic relationships within this diverse group remain obscure despite numerous studies of their evolutionary history using morphological, behavioral, ecological, and biochemical approaches (Duellman and Trueb, 1986; Cannatella and Hillis, 1993; Ford and Cannatella, 1993; Hedges and Maxson, 1993; Hillis *et al.*, 1993). Morphological traits are not always able to resolve branching patterns within the Anura largely because of the paucity of phylogenetically informative characters due to the strict conservation of the anuran body plan (Duellman and Trueb, 1986). Molecular methods that are useful for elucidation of specific and generic level relationships often are unable to resolve interfamilial relationships (Hillis, 1991; Maxson, 1992). Recently, DNA sequences of mitochondrial ribosomal RNA (rRNA) genes have been shown to be evolving at rates appropriate for resolving some aspects of amphibian phylogeny (e.g., Hedges and Maxson, 1993; Hay *et al.*, 1995). In addition, available fossil evidence indicates that major lineages of modern frogs diverged during a relatively short time span (Milner, 1988) which, if true, will complicate precise identification of relationships regardless of the method of phylogenetic inference employed.

The suborder Neobatrachia (Reig, 1958) was erected based on morphology and has recently received support from gene sequence data (Hedges and Maxson, 1993; Hillis *et al.*, 1993). The analysis of phylogenetic relationships among amphibian families based on more extensive mitochondrial rRNA gene sequences convincingly demonstrated the monophyly of this suborder (Hay *et al.*, 1995). The results of that study also corroborated the hypothesis of an early separation between the two major neobatrachian lineages: Bufonoidea and Ranoidea. However, only a few of the proposed relationships within either of these superfamilies were significantly supported.

One potential problem with most phylogenetic reconstructions is that of using only a single taxon to represent a large and diverse group. In such cases if the as-

sumption of the monophyletic origin of a group is not satisfied, the results of the analysis will be invalid. The impact of the species sampling scheme on the results of the phylogenetic analysis also can be significant (Lecointre *et al.*, 1993; Maxson *et al.*, unpublished data).

To improve the resolution of the neobatrachian portion of the anuran tree, we sequenced the same portions of the slowly evolving mitochondrial 12S and 16S rRNA genes that were used by Hay *et al.* (1995). The expanded data set from that study now includes representatives of all four currently recognized subfamilies of the Hylidae (Hemiphractinae, Hylinae, Pelodyadinae, and Phyllomedusinae), both subfamilies of the Myobatrachidae (Limnodynastinae and Myobatrachinae), two leptodactylid subfamilies (Leptodactylinae and Telmatobiinae), and three genera within the Dendrobatidae, a family traditionally associated with the ranoids (Duellman and Trueb, 1986; Ford, 1993), but according to the DNA sequence data, is nested within the Bufonoidea (Hedges and Maxson, 1993; Hay *et al.*, 1995).

In this study we address the following questions: (1) What are the relationships of the hylid subfamilies; in particular, what are the phylogenetic affinities of the Australian hylids (Pelodyadinae)? (2) Is the Myobatrachidae a monophyletic family and is it more closely related to the Heleophrynidae than to other bufonoid families? (3) Was the placement of the Dendrobatidae within the Bufonoidea (Hay *et al.*, 1995) an artifact of sampling or will its position remain the same with the inclusion in the analysis of additional members of the family? Finally, we anticipated that by including a wider representation of species, new patterns of relationships between different taxa might emerge.

MATERIALS AND METHODS

Taxa Examined

Eight new bufonoid species (see Appendix), representing the Dendrobatidae (*Colostethus*, *Phobobates*), Hylidae (*Gastrotheca*, *Litoria*, *Phyllomedusa*, *Smilisca*), Leptodactylidae (*Lithodytes*), and Myobatrachidae (*Pseudophryne*) were added to those species studied by Hay *et al.* (1995). All three mitochondrial ribosomal gene regions analyzed in this study were obtained from the same individual. Taxonomic assignment of examined species follows Duellman (1993).

DNA Amplification and Sequencing

Total DNA was extracted from frozen (-20°C) tissue samples of whole blood, plasma, heart, or muscle using a standard phenol-chloroform protocol (Hedges *et al.*, 1991) or Puregene DNA isolation kit (Gentra). A region of the 12S and two adjacent regions of the 16S mitochondrial rRNA genes were sequenced using the following scheme. Double-stranded fragments were amplified

in 30–35 cycles of the polymerase chain reaction (PCR; 95°C for 1 min, 50°C for 1 min, and 72°C for 2.5 min). Single-stranded sequencing template was produced in a second PCR amplification (25–35 cycles: 95°C for 1 min, 60°C for 1 min, 72°C for 2.5 min) with one primer as limiting, i.e., diluted to 1% of the original concentration (Gyllensten and Erlich, 1988). Slight variations in annealing temperatures and/or numbers of cycles were employed as needed to improve the quality of the DNA template. The primer pair 12L1/12H1 amplified a fragment of 12S rRNA gene (about 400 nucleotides long), while 16L1/16H1 (or 16L8/16H9) and 16L2a/16H10 yielded fragments of the 16S rRNA gene with approximate lengths of 500 and 570 nucleotides, respectively. Primers that were used for the double-stranded PCR were also used to generate a single-stranded template. In addition, a primer internal to the 16L2a/16H10 pair (16L2) was used. Location and sequences of all primers used were previously reported (Hedges, 1994), with the exception of 16H9 (5'-CCGGTCTGAACTCAGATCAGT-3'), which is colocalized with the 16H1 primer. The template was purified using 30,000 molecular weight filters (Millipore) and sequenced (Sanger *et al.*, 1977) using *Taq* (*Thermus aquaticus*) DNA polymerase with second round PCR primers as sequencing primers. A more detailed description of the methods has been published (Hedges *et al.*, 1991).

Sequence Analysis

Sequences were read from autoradiograms using the digitizing program GELIN (S. W. Schaeffer, Pennsylvania State University). We used only those portions of the new sequences which corresponded to the regions used in the analysis of amphibian phylogenetic relationships reported by Hay *et al.* (1995). The new data were combined with the sequences from representatives of 14 families of advanced frogs from that study. Since monophyly of both the Neobatrachia and the Archaeobatrachia is well established (Hay *et al.*, 1995) we used 4 representatives of the latter as outgroups in these analyses. The alignment was done by eye using the ESEE sequence editor (Cabot and Beckenbach, 1989). We omitted 359 sites from further analysis because of the moderate degree of length variation and presence of highly variable regions where homology could not be inferred with certainty.

Phylogenetic trees were constructed based on Jukes-Cantor corrected distances (Jukes and Cantor, 1969) using the neighbor-joining algorithm of Saitou and Nei (1987). We used the Jukes-Cantor distance in accordance with the recommendation of Kumar *et al.* (1993) since corrected distances were less than 0.3, average nucleotide frequencies did not deviate substantially from 25% (A, 32.7%; T, 22.6%; C, 23.7%; G, 21%), and transition/transversion ratios (range: 1–2.5, majority: 1.5–2) were not strongly biased. Nevertheless, for comparative purposes phylogenetic trees were constructed

using more sophisticated distance estimation methods that (a) account for deviation of nucleotide frequencies from 25% (Tajima and Nei, 1984), (b) consider differences in transitional versus transversional substitution rates (Kimura, 1980), and (c) both of the above conditions (Tamura and Nei, 1993). The resulting tree topologies were identical to the neighbor-joining tree reported here.

It has been shown that bootstrap confidence values tend to underestimate the extent of statistical support of species clusters, particularly when the true tree is starlike and the number of sequences involved in analysis is large (Sitnikova *et al.*, 1995). These circumstances are likely in our study since divergences within the Neobatrachia occurred during a short period of time (Milner, 1988), so we assessed statistical significance of groupings by the interior-branch test (Rzhetsky and Nei, 1992). Both the tree construction and statistical tests were done as implemented in the METREE program (Rzhetsky and Nei, 1994). Because the interior-branch test may overestimate statistical confidence when both the test and the tree reconstruction are done using the same data set (Sitnikova *et al.*, 1995), interior-branch confidence values (P_C) were corrected using a computer program provided by Tatyana Sitnikova (Pennsylvania State University). This correction gives a conservative estimate of statistical significance; therefore we treat values above 0.95 as significant and those above 0.90 as strongly supported. All calculations were done using the MEGA package (Kumar *et al.*, 1993).

Using MEGA, we also obtained maximum parsimony trees using the heuristic search method which does not guarantee that the tree(s) of minimal length will be found. The size of our data set prohibits the use of exhaustive or branch-and-bound searches, which provide such a guarantee. However, for this large data set, with long branches and short internodes, we believe the neighbor-joining algorithm, which has been shown to be more efficient in computer simulations (Nei, 1991; Nei *et al.*, 1995), is the most appropriate analytical method to use for recovering a phylogeny.

RESULTS

For each of the eight new taxa approximately 345 and 820 nucleotides were sequenced from the 12S and 16S rRNA genes, respectively. When combined with previously available data (Hay *et al.*, 1995) these sequences produced an alignment (Fig. 1) with a total length of 1258 nucleotides (376 for the 12S and 882 for the 16S). After conservative omission of sites where alignment was uncertain due to length and/or extensive sequence variation, a data set of 899 nucleotides was obtained (317 from the 12S and 582 from the 16S). Almost half (465) of those sites were variable; interestingly, the variation was equally distributed among both

genes (169 sites in the 12S and 296 sites in the 16S). This implies that within the Anura the requirements of sequence conservation due to structural constraints may be similar for these two genes.

The shaded areas in Fig. 1 were omitted from all analyses. Examination of the remaining alignment reveals that there still are some sites remaining which show a limited amount of length variation and missing data. There are two ways of analyzing our data set. All sites with gaps and missing data can be eliminated from the data set prior to the analysis (complete deletion) or they can be deleted from each pair of compared sequences separately (pairwise deletion). Since we had relatively few missing sites and they were distributed uniformly throughout the alignment we chose the latter option (Kumar *et al.*, 1993). When complete deletion was used the number of variable sites was reduced by about 10% (to 422 sites).

The phylogenetic tree reconstructed by the neighbor-joining method (Fig. 2) provides support for many groupings within the Neobatrachia. The monophyly of both the Bufonoidea ($P_C = 0.96$) and Ranoidea ($P_C = 0.78$) is supported. The three dendrobatid genera are monophyletic ($P_C = 0.98$) and cluster within the Bufonoidea. The two hylid subfamilies Pelodyadinae and Phyllomedusinae cluster together ($P_C = 0.90$), the Australian Myobatrachidae clusters with the Heleophryniidae ($P_C = 0.69$), the leptodactylid subfamily Telmatobiinae is a sister taxon of the New World bufonoids and their derivatives ($P_C = 0.92$), and there are several strong associations within the latter. Importantly, a neighbor-joining tree constructed using the complete deletion option (not shown) differed little from the tree presented here. Discrepancies between the two trees primarily involved a rearrangement of branches that received little statistical support, notably the relationships within the outgroup (Archaeobatrachia) and the position of the Sooglossidae (it was a sister group of the Telmatobiinae + neotropical Bufonoidea). In addition, a cluster containing the Centrolenidae, Hylinae, and Bufonidae with the Hemiphractinae was expanded in the "complete-deletion" tree to include the Leptodactylinae and Rhinodermatidae ($P_C = 0.95$).

The tree topologies derived from maximum parsimony analyses (with and without gaps and missing data) were fairly consistent with the neighbor-joining tree. All trees supported the division of the Neobatrachia into the Bufonoidea and Ranoidea and the placement of the Australian Myobatrachidae and the Heleophryniidae (although not as a monophyletic group) outside of the monophyletic Telmatobiinae + New World Bufonoidea. Also supported were the Hylinae (*Hyla* + *Smilisca*) and the Pelodyadinae + Phyllomedusinae joined consecutively by the Pseudidae and Rhinodermatidae. The most conspicuous differences between the neighbor-joining and the maximum parsimony trees involved the positioning of lineages with

	<12S	80
Discoglossidae	CCGCCAGGGAACCTACGAGCCTCAGCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Leiopelmatidae	CCGCCAGGGAACCTACGAGCCTCAGCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Pelobatidae	CCGCCTGGGAACCTACGAGCCTCAGCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Rhinophrynidae	CCGCCTGGGAACCTACGAGCCTAAGCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Bufonidae	TCGCCAGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Centrolenidae	TCGCCAGGGAACCTACGAGCAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Colostethus	TCGCCTGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Dendrobates	TCGCCTGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Heleophrynidae	TCGCCAGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Hemiphractinae	TCGCCAGGGAACCTACGAGCAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Hyla	TCGCCAGGGAACCTACGAGCAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Hyperoliidae	ACGCCAAGAACCTACGAGCGCAAGCTTAAAACCTAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Leptodactylinae	TCGCCAGGGAACCTACGAGCTAT - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Limnodynastinae	TCGCCAGGGAACCTACGAGCTAT - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Mantellidae	GCGCCAGGGAACCTACGAGCTAT - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Microhylidae	CCGCCAGGGAACCTACGAGCTAT - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Myobatrachinae	CCGCCAGGGAACCTACGAGCTAT - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Pelodyadinae	TCGCCAGGGAACCTACGAGCAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Phobobates	TCGCCAGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Phyllomedusinae	ANGCCAGAGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Pseudidae	TCGCCTGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Ranidae	GCGCCAGGGAACCTACGAGCTAT - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Rhinodermatidae	TCGCCAGGGAACCTACGAGCAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Smilisca	TCGCCAGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Sooglossidae	GCACCTGGGAACCTACGAGCTAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Telmatobiinae	TCGTCAGGGAACCTACGAGCAA - GCTTAAAACCCAAAGGACTTGGCGGTGCCNAAACCCACCTAGAGGAGCCTGTTCTA	
Discoglossidae	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - ACCGCCTATATACCACCGTCGCCAGCCACCCCGTGAG	
Leiopelmatidae	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - ACCGCCTATATACCACCGTCGCCAGCCACCCCGTGAG	
Pelobatidae	TAATCGATAATCCACGATCAACCTCACCATTCTTGCCAA - TCCGCCTATATACCACCGTCGCCAGCTTACCCTTTGAA	
Rhinophrynidae	TAATCGATAATCCCGCTTAAACCTCACCATTCTTGCCAA - ACCGCCTATATACCACCGTCGCCAGCCACCCCGTGAG	
Bufonidae	TAATCGATAATCCACGTTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Centrolenidae	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Colostethus	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Dendrobates	TAACCGATAATCCCGCTTAAACCTCACCATTCTTGCCAA - ACAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Heleophrynidae	TAATCGATGATCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Hemiphractinae	TCATCGATACTCCACGTTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Hyla	TAATCGATAATCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Hyperoliidae	TAATCGATAATCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Leptodactylinae	TAATCGATAATCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Limnodynastinae	TAATCGATGATCCACGTTTAAACCTCACCATTCTTGCCAA - CCCGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Mantellidae	TAATCGATAATCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Microhylidae	TAATCGATGATCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Myobatrachinae	TAATCGATGATCCCGCTTAAACCTCACCATTCTTGCCAA - CCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Pelodyadinae	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Phobobates	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Phyllomedusinae	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Pseudidae	TAATCGATAATCCCGCTTAAACCTCACCATTCTTGCCAA - TCCGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Ranidae	TAATCGATGATCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Rhinodermatidae	CAACCGATAACCCCGATTAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Smilisca	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Sooglossidae	TAACCGACACTACCAGATAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	
Telmatobiinae	TAATCGATAACCCCGCTTAAACCTCACCATTCTTGCCAA - TCAGCCTGTATACCTCCGTCGTCAGCTTACCACGTGAG	

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FIG. 1. Alignment of portions of mitochondrial 12S and 16S rRNA genes for all species in this study. “-” designates alignment gaps; “N” denotes undetermined nucleotides. Shaded areas were omitted from the phylogenetic analysis.

long branches (notably, the Sooglossidae and, to a lesser extent, Telmatobiinae and Hyperoliidae), an effect described by Felsenstein (1978). It should be reemphasized that because of the size of our data set (26 sequences of 899 nucleotides) we were unable to conduct a maximum parsimony search that guaranteed

finding the shortest tree (see Hedges *et al.* (1992) for discussion of a similar case). Furthermore, the nature of the tree (large number of taxa, long terminal branches, and short internal nodes) makes it difficult for a maximum parsimony algorithm to identify the correct tree (Nei *et al.*, 1995). Nonetheless, only nodes

Discoglossidae	GGGAATGCAGTAGGCAAAT-GGC	-----CCCCGC	CAGAACGTCAGGTCAAGGTGTA-GCGTATGAAGT
Leiopelmatidae	GGACTTCAAGTAGGCACAAT-GGT	-----CACCAC	CAAAACGTCAGGTCAAGGTGTA-GCACATGAAGT
Pelobatidae	GGTTAAAAAGTAAGTAAAT-GGT	-----ACTCAC	CAAAACGTCAGGTCAAGGTGTA-GCGAATGAAGT
Rhinophrynidae	-GCTGATTAGTAGGCTCAAT-GA	-----CACCAT	CAACACGTCAGGTCAAGGTGTA-GCGTATGGAGT
Bufo	CGCCAAATTAGTGAGCTTAAT-GTTT	-----TTTCA	CAACACGTCAGGTCAAGGTGTA-GCAAATGAAAT
Centrolenidae	CGCAATTTAGTGAGCTTAAT-GCCC	-----ATACGC	CAACACGTCAGGTCAAGGTGTA-GCAAATGAGAT
<i>Colostethus</i>	CGTC---AGTGAGCCCAAT-GTTT	-----GTTCAA	CCACACGTCAGGTCAAGGTGTA-ACATATAAAGT
<i>Dendrobates</i>	CGTCAGTGAGTGAGCCTAAT-GTTA	-----ATPCAA	CTTAAACGTCAGGTCAAGGTGTA-ACATATGTGAT
Heleophrynidae	CGTGAGARAAGTGGCCCTAAA-GAAC	-----CTTTT	CAATACGTCAGGTCAAGGTGTA-GCACATGAAGT
Hemiphractinae	CGACCCCCAGTGAGCTTAAT-GGCG	-----TTACAC	CAATACGTCAGGTCAAGGTGTA-GCAAATGAAAT
<i>Hyla</i>	CGTCACCCAGTGAGCCTAAT-GTTC	-----TTACAC	CAACACGTCAGGTCAAGGTGTA-GTAAATGAAAT
Hyperoliidae	TGTATT--AGTAAGTTAAATAGTAATAAC	-----AATTAC	CAAAACGTCAGGTCAAGGTGTA-GCCAACAAAATA
Leptodactylinae	CGTTTTATAGTGAGCTTAAC-GCTT	-----ATPCAC	CAGTACGTCAGGTCAAGGTGTA-GCTAATGAAAT
Limnodynastinae	CGGTCAATAGTGAGCAAAT-GGCC	-----AAGCGC	CAAAACGTCAGGTCAAGGTGTA-GCACATGAATA
Mantellidae	TGTAAGAAGAGTAGGTTTAAAG-GATC	-----CCCCAT	CAATACGTCAGGTCAAGGTGTA-GCCAAATGTAAT
Microhylidae	CGAATAA-AGTGAGCTTAAA-GGCC	-----COCGGC	CAACACGTCAGGTCAAGGTGTA-GTCCACGAAGC
Myobatrachinae	CGCCTAATAGTGAGCAAGAT-GGCC	-----TTTCGC	CAAAACGTCAGGTCAAGGTGTA-GCTAATGAAGC
Pelodyridinae	CGCAATCAGTAAGCTTAAT-GTCC	-----TTGGCT	CAATACGTCAGGTCAAGGTGTA-GTAAATAAAAT
<i>Phobobates</i>	CGTCAGTGAG---CTTAAT-GTTT	-----TTCCAA	CTATACGTCAGGTCAAGGTGTA-ACATATAAAAT
Phyllomedusinae	CGAACCCACAGTGAGCTTAAT-GCCA	-----GAACGC	CAATACGTCAGGTCAAGGTGTA-GCCAACAAAATG
Pseudidae	CGCTTTA-AGTGAGCCCAAT-GTCT	-----TCTCGT	CAATACGTCAGGTCAAGGTGTA-GTAAATAAAAT
Ranidae	CGTCTTC-AGTAGGCTCAAT-GATCATAA	-----TTACAT	CAATACGTCAGGTCAAGGTGTA-GCTTAAGAAAT
Rhinodermatidae	CGAAATATAGTGAGCTTAAT-GCCT	-----TTACAC	CAATACGTCAGGTCAAGGTGTA-GCAAATAAAAT
<i>Smilisca</i>	CGTTATTTAGTGAGCTTAAT-GTCT	-----CTACAT	CAATACGTCAGGTCAAGGTGTA-GTAAATAAAAT
Sooglossidae	TGAATTTTAAATAAGCCAAT-GGC	-----CCCCAC	CAATACGTCAGGTCAAGGTGTA-GCATATGTCTG
Telmatobiinae	TGAACCTTAGTAAGCCAAT-GTCTACACATAAATGGTTGAACAA	-----CAATAC	CAATACGTCAGGTCAAGGTGTA-GCGGATGATGT

Discoglossidae	GGAAAGAAATGGGCTACATTTTCTGGCT	-CAGAACAA	-CACGAAAGGTC	ACT-ATGAAA	-CCTGACC	--CAAAGGAGGA
Leiopelmatidae	GGGAAGAAATGGGCTACATTTTCTACCC	-TAGAACAA	-ACGAACGAT	CATT-ATGAAA	-CATGCTC	--AGAAGGTGGA
Pelobatidae	GGAAAGAAATGGGCTACATTTTCTAATAA	-CAGAATAT	-ACGAACGATT	ACT-ATGAAA	-AAGAATC	--TGAAGGAGGA
Rhinophrynidae	GGGAAGAAATGGGCTACATTTTCTAAAT	-TAGAAAAC	-AACGGAAGAC	CATT-ATGAAA	-CCTGGTC	--TGAAGGCGGA
Bufo	GGAAAGAGATGGGCTACACTTTCTAATT	-TAGAAAAAT	-ACGAAAAC	TACCTATGAAA	-CCTAGTT	--AGAAGGCGGA
Centrolenidae	GGGAAGAGATGGGCTACACTCTCTAATA	-TAGAAAAC	-ACGGAAGACT	TATGAAA	-CCTAGTC	--TAAAGGCGGA
<i>Colostethus</i>	GGAAAGAGATGGGCTACACTCTCTAACT	-TAGAACAA	-ACGAATAACT	TTA-ATGAAA	-CCTAACT	--AGAAGGCGGA
<i>Dendrobates</i>	GGGAAGAGATGGGCTACACTCTCTAACT	-TAGAATAT	-ACGAAAGACT	TATGAAA	-TCTAGTC	--AAAAGGTGGA
Heleophrynidae	GGAAAGAAATGGGCTACACTCTCTAACC	-TAGAAAAC	-ACGAAAGACT	GC-CTGAAA	-ACCAGTC	--TGAAGGCGGA
Hemiphractinae	GGGAAGAGATGGGCTACACTTTCTAART	-TAGAAAAA	-ACGAAAGACT	--TTATGAAA	-CCTAGTC	--GGAAGGCGGA
<i>Hyla</i>	GGGAAGAGATGGGCTACACTTTCTAATC	-TAGAACAC	-ACGAAAGACT	TATTTATGAAA	-CCTAGTC	--GGAAGGCGGA
Hyperoliidae	GGGAAGAAATGGGCTACAATTTCTACTT	-AAGAACAT	-ACGAATTTAAGT	AACTGAAA	-ACTACTA	--TGAAGGCGGA
Leptodactylinae	GGATAGAGATGGGCTACACTTTCTAACA	-TAGAATAT	-ACGAAAGACT	TACCTATGAAA	-TCTAATC	--TGAAGGCGGA
Limnodynastinae	GGGTAGAGATGGGCTACACTTTCTAART	-TAGAAAAAT	-ACGAAAAC	TACCTATGAAA	-TCTAGTT	--AGAAGGCGGA
Mantellidae	GGAAAGTAATGGGCTACAATTTCTATAA	-TAGAACAA	-ACGAAACACT	GC-ATGAAA	-AACAGTCA	--TGAAGGCGGA
Microhylidae	GGAAAGAAATGGGCTACAATCTCTATCT	-TAGAACAA	-ACGAAAGACC	CAC-ATGAAA	-AACAGTCA	--TGAAGGCGGA
Myobatrachinae	GGGCAGAGATGGGCTACATTTTCTACTC	-TAGAAAACC	-ACGAAAGACT	GC-GTGAAA	-CCCCGTC	--AGAAGGAGGA
Pelodyridinae	GGGAAGAGATGGGCTACACTTTCTACCT	-TAGAATAA	-ACGAAAAC	TATTTATGAAA	-TCTAGTT	--AGAAGGCGGA
<i>Phobobates</i>	GGAAAGAGATGGGCTACACTTTCTACCA	-TAGAATAA	-ACGAAAGACT	TA-ATGAAA	-CCTAGTC	--AGAAGGCGGA
Phyllomedusinae	TGGAAGAGATGGGCTACACTCTCTAART	-TAGAACAT	-ACGAAAAC	TTT-ATGAAA	-CCTAGTT	--AGAAGGCGGA
Pseudidae	GGGAAGAGATGGGCTACACTCTCTACTA	-TAGAGTAA	-ACGAAAGACC	CCTATGAAA	-CCTGGTC	--AG-AGGCGGA
Ranidae	GGGAAGCAATGGGCTACAATTTCTAATA	-TAGAACAA	-ACGAAAGGCT	AT-GTGAAA	-TCATAGCAGC	--GAAGGCGGA
Rhinodermatidae	GGGAAGAGATGGGCTACACTTTCTAACA	-TAGAAAAT	-ACGAAAGACT	TACATATGAAA	-CCTAGTC	--AGAAGGCGGA
<i>Smilisca</i>	GGGAAGAGATGGGCTACACTCTCTAACT	-TAGAGTAC	-ACGAATGACT	TATTTATGAAA	-TCTAGTC	--AGGGGCGGA
Sooglossidae	GGGCAGAAATGGGCTACACTCCCTCACAA	-CAGGAAAAT	-ACAAAAGACC	-TAGAGAAA	-CAAAGTC	--AAAAGGCGGA
Telmatobiinae	GGAAATGTGATTGGGCTACACTTTTCTAART	-TAAAAGAA	-ACGGAAGACT	AAAAATGAAA	-CTCTGTC	--AGAAGGCGGA

FIG. 1—Continued

poorly supported on the neighbor-joining tree varied in their position in the maximum parsimony trees.

DISCUSSION

The addition of eight new sequences representing members of different bufonoid lineages and using a

phylogenetically closer outgroup, the Archaeobatrachia, has allowed us to clarify some relationships with the Neobatrachia. There is support for a split at the base of the radiation between the two major groups (Bufonoidea and Ranoidea) within the suborder. We now can securely place the Sooglossidae in the Bufonoidea ($P_c = 0.96$). Previous morphological studies had

Discoglossidae	TTTAGTAGTAAAGAAAAA	CAGAGATTCTCTTTAAC	TCGGCCCTGGG-CGCGCCACAGCTCATGCAAGCTTAACCA
Leiopelmatidae	TTTAGCAGTAAAAAGGAACA	AAGAGTCCCTTTTAAAT	CCGGCCCTGGG-CGCGTCACAGCTTNGCACCCCTAACCC
Pelobatidae	TTTAGCAGTAAAAAGGATCA	TGATGCCCTTTTAAAT	CTGGCCCTGGG-CGCGCCACAGCTCCCCCTCAATTAACCTT
Rhinophrynidae	TTTAGCAGTAAAAAGAAACAG	GAGAGTCTTTTAAAGG	CGGCCCTGGGCGCGCAAGCT--ATCTACCATAAACCAA
Bufo	TTTAGCAGTAAAAAGGAATCA	GCATGTCCATTTTAAAC	CGCACACTGGG-TGTGTAAAGCTTAATTTGTTTAAACGCT
Centrolenidae	TTTAGAAGTAAAAAAAATAT	GAATGTCCCTTTTAAAC	TGGGCACTGGGGTGTGTCAAAGCTTAATCNANCCACTAAT
Colostethus	TTTAGCAGTAAAAAGAACTA	GAGCGTTCATTTAAAT	TAGGCACTGGGGTGTGTCAAAGCTTCCTCATCTGTTCTGCCC
Dendrobates	TTTAGAAGTAAAAACGTAACAA	GAGAGTCCCTTTTAAAC	ATGGCACTGGG-TGTGTCAAAGCTTTCTTACACCCAAAGAT
Heleophrynidae	TTTAGTAGTAAAAAGAAACAA	GAGAGTCTCTTTTAAAC	TCGGCCCTGGGGTGTGTCAAAGCTTCACCATCTACACCCAA
Hemiphractinae	TTTAGAAGTAAAGGGGAAATCA	GAATGTCCCNNTTAAAC	TTG-CACTGGGGTNTGTCAAAGCTTATTCTGACTCTTATTT
Hyla	TTTAGAAGTAAAAAAGATCA	TAAAACCTCTTTTAAAC	CCGGCACTGGGGTGTGTCAAAGCTTCATTTATCCCGCGCTCC
Hyperoliidae	TTTAGAAGTAAAAAGAAACCA	AAGTGTCTTTTAAAC	TCGGCACTGGGGTGTGTCAAAGCTTCACCTCGCAACTCTGTAT
Leptodactylinae	TTTAGCAGTAAAAAGGAAACA	GAAAGTCCCTTTTAAAC	ACGGCCAGGG-GATGTATAGCTTAAACACACCCCTCAACA
Limnodynastinae	TTTAGTAGTAAAAAGGGGATA	GAGAGTCTCTTTTAAAC	AGGCCCTGGGACGTGTNAAAGCTTAAACATTTTCTTTTCT
Mantelliidae	TTTAGTAGTAAAAAGAAATA	AAGTGTCTTTTAAAT	TAGGCCCTGGGACGCGTCAAAGCTTAAATACATAATAAATA
Microhylidae	TTTAGTAGTAAAAAGGAAATCA	GAGAGTCTCTTTTAAAC	CCGGCACTGGGGTGTGTCAAAGCTTTTACATCTCATCAACT
Myobatrachinae	TTTAGTAGTAAAAAGAAATCA	GAGAGTCTCTTTTAAAC	CCGGCACTGGGGTGTGTCAAAGCTTAAATTTTAAATAA
Pelodryadinae	TTTAGCAGTAAAGGTGAAACTA	GAGCGTTCATTTAAAT	ACGGCACTGGGGTGTGTCAAAGCTTCCTCATTTTCTCTCC
Phobobates	TTTAGCAGTAAAAAGAACTA	GAGAGTCTCTTTTAAAT	AAGCACTGGGGTGTGTCAAAGCTTAAATTTTAAATAACTC
Phyllomedusinae	TTTAGCAGTAAAAAGGAAACA	GAGAGTCTCTTTTAAAC	CTGGCACTGGGGTGTGTCAAAGCTTACTTGTGTATAACATTT
Pseudidae	TTTAGTAGTAAAAAGAAATA	GAGTGTCTCTTTTAAAC	CCGGCTCTGGGATGCGTNNAGCTTAGTCTTACACACTTAC
Ranidae	TTTAGCAGTAAAAAGATAACA	GAGAGTCTCTTTTAAAT	TTGGCACTGGGGTGTGTCAAAGCTTAAATTTCTTATTATTA
Rhinodermatidae	TTTAGGAGTAAAAAGAGACA	GAGAGTCTCTTTTAAAC	TTGGCACTGGG-TGTGTCAAAGCTTCATTTTCTTTCTTT
Smilisca	TTTAGTAGTAAAAAGAAACA	GAGTGTCTCTTTTAAAC	TCGGCCCTGGGACAGCTCACAGCTTACCCTTCAAACCCAAA
Sooglossidae	TTTAGAAGTAAAGAAAGATA	TCATATTTTCTTAAAT	AAGGCACTGGAGTATGTNNMNTTAGCTATTATACTTAAAC

Discoglossidae	A--CAATACCGTTTAAACCC	TCTAAACCCATAAATACTAATGAGCCTCTCTATAATTT	TATAGAACAGTTTATGCTAG
Leiopelmatidae	CAATAAT-ACGAATAACCAATCAAAACCCATCAACACTACTGGCCCTATCTATAAACT	CATGGATACGCTTATGCTGG	
Pelobatidae	TT--AAT-TAGGACNCAAAATTCAAAACCCATAAAACTTATGAGTATTCCTATACGTT	TATAGGAGAATTAATGCTAG	
Rhinophrynidae	C---AAT---TTAAACCTAATCAAACCCCTAACACACTACTGAGTGTCTATACACC	TATAGAACCCTTATGCTAG	
Bufo	---AATTACTAAACTTCCATCTTAATCTTCACTAATACTGAGTAATTTCTATAATTTCT	TATAGAAATTTATGTTAA	
Centrolenidae	---AAT-CCCAACATTAAGAACCCCTTACATAGTACTGAGTAATTTCTATAATCAC	TATAGAAGACTTTATGTTAA	
Colostethus	T--AATTCCTCTACCTAT-CATTAACCCCTTCCCCCAAAGAGCAATTTCTATAATGTT	TATAGAACTATTATGTTAA	
Dendrobates	---AATTCGAANACCTTTCTCTACCCCTTACCCCTTATAGAGCAACTCTATACTTC	TATAGAACTTTTATGTTAA	
Heleophrynidae	T--AAT-CCCACAACATCTCAAACCCCTCACCCATCAAGCAATTTCTATAATCCC	TATAGAACATCTTATGCTAG	
Hemiphractinae	TA--AATTTCTCACCATT-AATTAACCCCTTCAAACAATACTGAGTAATTTCTATAATGTC	TATAGAAATTTTATGTTAA	
Hyla	C--AAT-ACCTCAATGATCCAGAACCCTTCTCCTCACTAATAAGTACTTCTATAATCCC	TATAGAACATTTTATGTTAA	
Hyperoliidae	A--AATTTTAAATTTGACTATAAAACCCCATCAATAACCAATAACTTTATAATTA	TATAAAAAATTTATGTTAG	
Leptodactylinae	T--AATACCCTACTAA-AGAGAAACCCACTTCAATATTAAGCAATCCATAATCCC	TATAGNAGGTTTATGTTAA	
Limnodynastinae	TC--AAT-ACCACCATCCCAACAACCCCTTCTCCAGTATCAAGCAATTTCTATAGAGA	TATAGAAAACTTATGCTAG	
Mantelliidae	---AAT-TCTCAAAACCCCTTAAACCCCTCCCAATATTGAGTAATTTCTATAAATA	TATGGAAGATATTATGCTAG	
Microhylidae	---AAT-ACCAAAACACTTACATAACCCCTCACCCAGTACTAAATAACTCTATAATCCC	TATAGAAATTTATGCTAA	
Myobatrachinae	A--AATACCCTCCACCGA-TCRAAACCACACACTTATGAGTGTATTTCTATAATTT	TATGGAAGACTTTATGCTAG	
Pelodryadinae	---AATTCAAAAATTTAT-AAAAACCCCTTAACTACTATTAAGTACTTCCATAATCTT	TATGGAATTTACAAATGTTAA	
Phobobates	---AATTCCTCTCTTTT-TAACAACCCATTTTCCAGAGAGCAATTTCTATAATTA	TATAGAAATTTTATGTTAA	
Phyllomedusinae	---AATCCCTCTAGTTAT-TAAAACCCCTTAACTAATTAAGTAAATTTATAAATA	TATAAAACATATAATGTTAA	
Pseudidae	TTTCAAT-ACCCTAATTTTCCAGACCCCTTTATTTTATGAGTGTCTATAATTA	TATAGAAATTTATGTTAA	
Ranidae	---AAT-TTCTAAATTTTATGATCAACCCCTTACTGAAATTTTATAATCCC	TATAAAAAGTAATAATGCTAG	
Rhinodermatidae	---AAT-TTCTTCAATCAATTAATACCCCTTAAATAGTACTGAGCACTTCTATAATTT	TATAGAAATTTTATGTTAA	
Smilisca	---AAT-ACCACAATTTATTTAGAACCCCTTCAATATTGAGAACTTCTATAATCCC	TATAGAAATTTTATGTTAA	
Sooglossidae	---AAT-CCCCTCAACACTACAGAACCCTGATACACCATTAAGCAATCTATTTCCCA	TAT-GAAC-TATTATGCTAA	
Telmatobiinae	---AATTTCTAAATAAATTAACCCCTACCTTAATACTGAATAATTTCCATAATA	TATGGAAGCTATTATGTTAA	

FIG. 1—Continued

alternately located this family in the Bufoidea (Laurent, 1979), or the Ranoidae (Duellman, 1975). Placement of Sooglossidae close to the Myobatrachinae (Ford and Cannatella, 1993; Lynch, 1973) within the Bufoidea remains a possibility, since our phylogenetic tree does not definitively identify a sister-group of the Sooglossidae.

The Heleophrynidae is still most closely associated

with the Australian Myobatrachidae (Limnodynastinae) as found by Hay *et al.* (1995). However, inclusion of representatives from both myobatrachid subfamilies suggests that the Myobatrachidae may not be monophyletic. In their review of phylogenetic relationships among major frog lineages, Ford and Cannatella (1993) found evidence for the monophyly of the Limnodynastinae and of the Myobatrachinae, but not of the family

Discoglossidae	AACTAGTAAC-ACGAAATA	ATTCTCT	--AAATGCAAGTGTAAATCAGATCGAAAACCTCACTGATAATNAACGTAC	560
Leiopelmatidae	AATGAGTAAC-AAGAATTAAC	CATCTCC	--CAATGTAAGTGCACCAGACCGAATCAATCACTGGCAATTAACGGCC	
Pelobatidae	AACTAGTAAC-TAGAATTAAC	ATTCTCT	--TAATGTAAGTGTAAATCAGATCGAATAAATCACTGATAATTAACGGCC	
Rhinophrynidae	AACTAGTAAC-TAGAATTT	ATTCTCC	--AAATGCAAGTGTAAATCAGATCGAAAACCAATCACTGATAATTAACGACC	
Bufo	AACTAGTAAT-AAGAAGAGAT	CTTCTCT	--TAATGTAAGTGTAAATCAGAAAGGACAAACCCTGATATTTAACGTCA	
Centrolenidae	AACTAGTAAT-AAGAAGAGAA	CTTCTCT	--ACATGCAAGTGTAAATCAGATAGGACACTCCACTGATAATTAACACCA	
Colostethus	AACTAGTAAT-AAGAAGTAGAA	CTTCTCT	--TAATGCAAACTACATCAGCAAGGACACCCCACTGATTCTTATCGCCC	
Dendrobates	AACTAGTAAT-AAGAAGTAGAA	CTTCTCT	--AAAGCAAGTTTGCCTCAGTTCCGACACCCCCCTGACAATTAACGTTT	
Heleophrynidae	AACTAGTAAC-AAGAAGTAGAA	CTTCTCCA	--CAATACACGTGTAAATCAGATTGGACCACCCCTGATAATTAACGCCA	
Hemiphractinae	AACTAGTAAC-AAGAAGTAGCC	CTTCTCT	--AAATGCGCCCGTAAATCAGAAAGGACCCCCCACTGATAATTAACGCCCT	
Hyla	AACTAGTAAC-AAGAAGTAGAA	CTTCACT	--AAATGCAAGTGTAAATCAGAAAGGACCTACCCTGACTTTTATCATAC	
Hyperoliidae	AATGAGTAAT-AAGAAGTAGAA	TTTCTCCA	--AAATATAAGCATAATGCAAAATGAACTAATCACTGGCAATTAACGTAT	
Leptodactylinae	AACTAGTAAC-AAGAAGTAGAA	CTTCTCAT	--TAGTGCATGTGTAAATCAGACAGTACATTCCTGATCCCTAACACTA	
Limnodynastinae	AACTAGTAAC-AAGAAATAGAC	CTTCTCT	--CAATATAAGTGTACATCAAAATGGACACACCCTGATAACTAACGCCA	
Mantellidae	AACTAGTAAC-AAGAAATTAAC	TTTCTCCA	--AAATGTAAGTGTAAAGCCACATAGATAAACTAATGGCAATTAACGAAA	
Microhylidae	AACTAGTAAC-ATGAAGTAGCC	CTTCTCT	--AAATGTAAGTGTAAAGCCAAAAGAACCAATCACTGGCAATTAACGTAT	
Myobatrachinae	AATTAGTAAT-CCGAAGTAGCC	CTTCTCT	--CAACACAAGTGTAAATCAGAAAGGACACCCCCCTGATAATTAACGCC	
Pelodyadinae	AACTAGTAAC-AAGAAGTAGCC	CTTCTCT	--AAATGCAAGTGTAAATCAGAAAGGACACACCCTGATAATTAACGCCCT	
Phobobates	AACTAGTAAT-AAGAAGTAGCC	CTTCTCT	--TAATGCAAACTTAAATCAGTCCAGGACCATCCGCTGATCTTTAACGCTT	
Phyllomedusinae	AACTAGTAAC-AAGAAGTAGCC	CTTCTCT	--AAATGCAAGTGTATGTCAGATTGGACAAACCCTGACTTTAACCATCT	
Pseudidae	AACTAGTAAC-AAGAAGTAGCC	CTTCTCT	--AAATGCGAGTGTATATCAGAAAGGACAAACCCTGATAATTAACGCC	
Ranidae	AACTAGTAAC-AAGAACTGCCC	ATTCTCT	--AAATGCAAGCATAAAACAGAAATAGACACCCTACTGGTAATCAACGTAA	
Rhinodermatidae	AACTAGTAAC-AAGAAGTAGCC	CTTCTCT	--AAATGCAAGTGTAAATCAGAAAGGACACACCCTGCTAATTAACAGTC	
Smilisca	AACTAGTAAC-AAGAAGTAGCC	CTTCTCT	--AAATGTAAGTGTAAATCAGAAAGGACACTACCCTGACTTTAACCATCT	
Sooglossidae	AACTAGTAAT-AAGGGATCAA	CCCCTCT	--TAATGCAAACTACATCAGCACGGATAAAACCACCGATACTTTAACGCC	
Telmatobiinae	AACAAGTAAC-ATGAAGTAGCC	CTTCTCT	--AAATGTAACCTAAGTCCAGACCCGATAAAACACTGACAATTAACAGCC	

Discoglossidae	CACA-TGAAGAACTGTAGTAACCTAT	CAAGAAAA	--TCCTAC-AACCGCCACCGTTAATCTAACACAAGAGCAT	640
Leiopelmatidae	TCAA-TGAGACAAAATGTAGCAACTCCA	CAAGAAAA	--CCCTAC-AACCACCACCGTTAACCTAACACAAGAACAT	
Pelobatidae	TTAC-AGAAAGCAATGTAAACAACCTAC	CAAGAAAA	--CCTTAC-ATCAAAAATGTAAATCTTACACAAGAACAC	
Rhinophrynidae	TTAAATGAAAAAATGTAGCAACCAC	CAAGAAAA	--CCCTAT-ACCACACACCGTTAACTTACACAAGAGCAT	
Bufo	C---TGAGTTAAAAGTAAAAACTTAT	CAAGAAAA	--CTCTAC-TTCCATGAAGCTTAACTTAACTAACACAAGAGCAT	
Centrolenidae	A---TGAAACAAAAGTAGTAACCTCAA	CAAGAAAA	--CTCTAC-TACATTTTTATGTTAATCTAACACCAGAGCAT	
Colostethus	T---TAAAAACATGGCAGCAACTTAT	CAAGAAAA	--CACTCC-CTCCATCAGCGTTAACCTAACACCAGAGCAT	
Dendrobates	T---TAATTTTAAAGTAGTAACCTTAA	CAAGAAAA	--GCCTAC-TAAAACCTTACGTTTACCTTACACTAGCGCGT	
Heleophrynidae	---TGAACTTAGGGTAAATAACGACA	CAAGAAAA	--ACCTACCACCCCGCGTTAACCTAACACAAGAGCAT	
Hemiphractinae	A---TGACCCTAAAAGTAAAAACTTAT	CTAGAAAA	--ACTTAC-TAAAGCGCCGTTAACTTAACTAACACAAGAGCAT	
Hyla	A---TGCAAAACAAATTAATAAATTTTT	CAAGAAAA	--ACTTAA-TTTTAACTATGTTAATCTAACACAAGAGCAT	
Hyperoliidae	A---TTAACTTAAAGTAGTAACCTTAA	CTAGAAAA	--CCCTAC-TTTTCTAAACATTAATCTAACACAAGAGAT	
Leptodactylinae	T---TGAATCTTAA-TAGCAACTTTA	CAAGAAAA	--CCCTAT-TATTTTATATGTTAATCTAACACAAGAGCAT	
Limnodynastinae	C---TGAAACCCATAGTAGCAACCAC	CAAGAAAA	--CCCTAC-TAACCCCAACGTTAACCTAACACTAGAGTAT	
Mantellidae	A---TGAAACCCCTTTAGTAACCTAAT	CAAGAAAA	--TTCTAC-AAACCAAAACGTTAACCTTACACTAGAACAT	
Microhylidae	C---TGACCCCTTAAAGCAACAACCTATA	CAAGAAAA	--TCTTGC-TAACACCAACGTTAACCTAACACAAGAGCAT	
Myobatrachinae	A---TGAAAAAAGAACAGTAACTTAA	CTAGAAAA	--TCCTGC-TCATTAATCCGTTAACCTAACACAAGAGTGT	
Pelodyadinae	A---TGAACTCAAAGTAGTAACCTTCA	CAAGAAAA	--TTCTAC-TAAATCTAATGTTAACCTAACACAAGAGCAT	
Phobobates	T---TAAACCACCTGTAGTAACCCAT	CAAGAAAA	--CACTAC-CTCACTTAGCGTTAACCTGACACAAGAGCAT	
Phyllomedusinae	---TGAAATCAAAGTAAACAACCTTAT	CAAGAAAA	--TTTTCAC-CAAAACAGCTGTTAATCTAACACAAGAGCAT	
Pseudidae	A---TAAATAATAGTAAATAACATAA	CAAGAAAA	--CCTTAC-TCTCTTACAGCTTAACTTAACTAACACAAGAGCAT	
Ranidae	A---TGTCACCTTTTATAGTAACCTAG	TAGAAAA	--TCCTAATCCCTTACGTTAACCTTACACTAGAGCAT	
Rhinodermatidae	C---TGAATATAAAGTAGTAACCCAA	CAAGAAAA	--TTCAAC-TAAATCTCATGTTAACCTAACACAAGAGCAT	
Smilisca	A---TGAAAATACATAGCAACTCCA	CAAGAAAA	--ACCTAA-TTGTAAACCATGTTAACCTAACACCAGAGCAT	
Sooglossidae	AC---TAATCCCGCGCTACACAAAACACA	CAAGAAAA	--TTTACC-TAAGCCAAACGTTAACCTAACACAAGAGCAT	
Telmatobiinae	---TTAACTAAAAGTAGTAACCCAA	CAAGAAAA	--TCTCACT-TAACCTACTGTTAACCTAACACAAGAGCAT	

FIG. 1—Continued

Myobatrachidae, which consists of both lineages. Expanded sampling of members within both of these old lineages (Maxson, 1992) will be necessary to identify the sister group of the South African Heleophrynidae.

The position of the Dendrobatidae within the Neobatrachia has elicited much controversy. It was previously placed both within the Ranoidea (Duellman and Trueb, 1986; Ford, 1993) and the Bufonoidea (Lynch,

1973). With the addition to the analysis of two more dendrobatid genera we substantiated the placement of the Dendrobatidae within the bufonoids. Relationships within this family will be discussed in more detail elsewhere (I. Ruvinsky, B. J. Smith, and L. R. Maxson, unpublished data).

Monophyly of a bufonoid subgroup (containing all taxa except the Heleophrynidae, Myobatrachidae,

Discoglossidae	AA - AAGAAAGATTTAAAAGATATAAAAAGGAAC TCGGCAAACATGAAC TCGCCTGTTTACCAAAAAACATCGCCTCTGCTC
Leiopelmatidae	ACTAGGAAAAGATTTAAAAACAAGGAAGGAAC TCGGCAAATATGAACCTCGCCTGTTTNCACAAAAACACCGCCTCTTGCAA
Pelobatidae	ACCAAGAAAGATTTAAAAGATATAGAAGGAAC TCGGCAAATCTAGGTCCCGCCTGTTTACCAAAAAACATCGCCTCTTGCCC
Rhinophryniidae	TCCAGGAAAAGATTTAAAAGACTTGGGAAGGAAC TCGGCAAACAAGAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGCCC
Bufo	TTCAGAAAAGATTTAAAAGAAAAAGAAGGAAC TCGGCAAATATTAACCTCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Centrolenidae	TACAAGAAAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATATTAACCTCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Colostethus	TTTAAGAAAAGATTTAAAAGAAAAAGAAGGAAC TCGGCAAATAATAGCCCCGCTGTTTACCAAAAAACATCGCCTCTTGC--
Dendrobates	TACAAGAAAAGATTTAAAAGAAAAAGAAGGAAC TCGGCAAATATTAGTCTCGCCTGTTTACCAAAAAACATCGCCTCTTGC--
Heleophryniidae	TTCTGGAAAAGATCTAAAAGGAGAGGAAGGAAC TCGGCAAATTTTGGCCCCGCTGTTTACCAAAAAACATCGCCTCTTGAAT
Hemiphraclinidae	TTACAAGAAAAGATTTAAAAGAAAAAGAAGGAAC TCGGCAAATATTAGCCCCGCTGTTTACCAAAAAACATCGCCTCTTGA--
Hyla	TTCAAGTACGATTTAACGAAAAAGAAGGAAC TCGGCAAATATTAACCTCGCCTGTTTACCAAAAAACATCGCCTCTTGC--
Hyperoliidae	TACTGGAAAAGATTTAAAAGAAATGGGAAGGAAC TCGGCAAATATAAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGC--
Leptodactylinae	TTACAAGAAAAGATTTAAAAGAAAAAGAAGGAAC TCGGCAAATCTTAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Limnodynastinae	TTTGAGAAAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATATTAACCTCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Mantellidae	TAATGGAAAAGATTTAAAAGAAAAAGAAGGAAC TCGGCAAATCTTAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Microhylidae	TTCAGGAAAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATTTTAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Myobatrachinae	TAATAGAAAAGATTTAAAAGGAGCGGAAGGAAC TCGGCAAATACCAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Pelodryadinae	TAAAAGAAAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATATTAACCTCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Phobobates	TCCAAGAAAAGATTTAAAAGAAAAAGAAGGAAC TCGGCAAACACCCAGCCCCGCTGTTTACCAAAAAACATCGCCTCTTGT--
Phyllomedusinae	TTTCAGACAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATATTAACCTCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Pseudidae	TACAAGAAAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATATTAAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Ranidae	TTCAAGAAAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATTTAGCCTCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Rhinodermatidae	TACAAGAAAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATATTAAACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Smilisca	TTCAAGTAAGATTTAAAAGAAAAGAAGGAAC TCGGCAAATATTAACCTCGCCTGTTTACCAAAAAACATCGCCTCTTGA--
Sooglossidae	TACAAGAAAAGATTTACAAGAAAAGAAGGAAC TCGGCAAATTTTACCCCGCCTGTTTACCAAAAAACATCGCCTCTTGCCT
Telmatobiinae	TTTAAGAAAAGACTTAAAGAGAAAAGAAGGAAC TCGGCAAATAATTGATTCGCTGTTTACCAAAAAACATCGCCTCTTGA--

Discoglossidae	T - - - ACATGTATAGGAGGTCCAGCCTGCCAGTGACGTAAATGTTTAAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNNCCT	16s2>16s1	800
Leiopelmatidae	ACAAACATATATAAGAGGTATAGCCTGCCAGTGACAATT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNNACC		
Pelobatidae	AR - - - TCTGTATAAGAGGTCCCGCCTGCCAGTGACATCA - - - GTTAAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Rhinophryniidae	AA - - - TTCAAATAAGAGGTCCAGCCTGCCAGTGACCATG - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Bufo	- - - ATACTCATAAGAGGTCCAGCCTGCCAGTGACCAT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Centrolenidae	- - - ATAAATATAAGAGGTCCAGCCTGCCAGTGACTCT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Colostethus	- - - - - CACCTATAAGAGGTCCAGCCTGCCAGTGACTCT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Dendrobates	- - - - - AAACAATAAGAGGTCCAGCCTGCC - - - AGTGACTCATT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Heleophryniidae	- - - - - ACATAAAGAGGTCCAGCCTGCCAGTGACTTT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Hemiphraclinidae	- - - - - AAAACATAAGAGGTCCAGCCTGCCAGTGACTACT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Hyla	- - - - - AAAATATAAGAGGTCCAGCCTGCCAGTGACTTT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Hyperoliidae	- - - - - AAATATAAGAGGTCCCGCCTGCCCGTGATTTA - - - GTTTAAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Leptodactylinae	- - - - - CAACTATAAGAGGTCCAGCCTGCCAGTGACTCT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Limnodynastinae	- - - - - TTATATAAGAGGTCCAGCCTGCCAGTGACCAT - - - GTTGANNNNNNNNNNNNNNNNNNNGTCT - - - TAAT - AGGACT		
Mantellidae	T - - - - CAAATATAAGAGGTCTAGCCTGCCAGTGACAA - - - ATTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Microhylidae	- - - - - ACATATAGAGGTCCAGCCTGCCAGTGACCAA - - - GTTAAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Myobatrachinae	- - - - - CAAATATAAGAGGTCCAGCCTGCCAGTGACTTA - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Pelodryadinae	- - - - - ACCATAAGAGGTCCAGCCTGCCAGTGACTCT - - - GTTAAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Phobobates	- - - - - CAATTATAAGAGGTCCAGCCTGCCAGTGACTTT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Phyllomedusinae	- - - - - AACATATAAGAGGTCTAGCCTGCCAGTGACTCT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Pseudidae	- - - - - TATTATAAGAGGTCCCGCCTGCCAGTGACTCT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Ranidae	- - - - - AAAATATAAGAGGTCCAGCCTGCCAGTGACTAA - - - GTTAAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Rhinodermatidae	- - - - - AACATAAGAGGTCCAGCCTGCCAGTGACTATA - - - ATTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Smilisca	- - - - - AAAATATAAGAGGTCCAGCCTGCCAGTGACTCT - - - GTTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Sooglossidae	- - - - - AAACATAGAGGTCTGCTGCCAGTGACTCA - - - ATTCAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		
Telmatobiinae	- - - - - AAATCATAAGAGGTCCAGCCTGCCAGTGACTCT - - - GTTAAACGGCCGCGGTATTNNNNNNNNNNNNNNNNNNN		

FIG. 1—Continued

Sooglossidae, and Telmatobiinae) is strongly supported, with the Telmatobiinae strongly supported as the sister taxon to this subgroup. These findings provide additional support for the bufonoid phylogeny reconstructed with less extensive species sampling (Hay *et al.*, 1995). Identification of a well-supported pair consisting of the Pelodryadinae and Phyllomedusinae provides a basis for some of the following discussion where

we address evolutionary and biogeographic implications of the expanded phylogenetic scheme for the Bufo-noidea.

Members of the Sooglossidae are restricted to two islands of the Seychelles Archipelago which has been separated from the Indian continent for over 60 million years (Dickin *et al.*, 1986/1987). It can be proposed that the ancestor of the Sooglossidae was separated from the

Discoglossidae	AGTATGAATNGCACCACGAAAGTTCACTGTCTCCTATATCTAATCAGTGAACTGATCTCCCCGTGAGAAAGCGGGGAT
Leiopelmatidae	AGTATGAACGGCACCACGAGGGCTCACTGTCTCCCTGTTAATCAGTGAAATGATTTCCAGTGCAGAAAGCTGGGAT
Pelobatidae	GGTATGAACGGCACCACGAAAGCTTACTGTCTCCTATATCCAATCAGTGAACTGATCTCCCCGTGAGAAAGCGGGGAT
Rhinophrynidae	GGTATGAATGGCACCACGAAAGTTACTGTCTCCTTTTCTAATCAATGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Bufoidea	AGTATGAATGGCACCACGAAAGTTACTGTCTCCTTTTCTAATCAATGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Centrolenidae	TGTATGAATGGCTCCACGAGGGTTACTGTCTCCTTTTCCAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
<i>Colostethus</i>	AGTATGAATGGNCCACGAGCGGTACGTGTCTCCTTTTCTAATCAATGAACTAATCTTCTCGTGAAGAAGCGGAGAT
<i>Dendrobates</i>	AGTATGAACGGCTTACGAAAGACTACTGTCTCCTTTTCTAATCAATGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Heleophrynidae	TGTATGAATGGCTCCACGAGGGTTACTGTCTCCTTTTCCAATCAGTGAACTGATCTCCCCGTGAGAAAGCGGGGAT
Hemiphractinae	NNNNNNNNNNNACCACGAGGGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
<i>Hyla</i>	AGTATGAATGGCACCACGAAAGTTACTGTCTCCTTTTCCAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Hyperoliidae	NNNNNNNNNNNNNACGAGGGTTACTGTCTCCTTTTCTAATCAATGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Leptodactylinae	NNNNNNNNNNNNNCCGAGGGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Limnodynastinae	AGTATGAATGGCACCACGAGGGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Mantellidae	NNNNNNNNNNNNNNNCCGAGGGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Microhylidae	AGTATGAACGGCACCACGAAAGTTACTGTCTCCTTTTCCAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Myobatrachinae	AGTATGAATGGCACCACGAGGGTTACTGTCTCCTTTTCCAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Pelodyadinae	AGTATGAATGGCACCACGAAAGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
<i>Phobobates</i>	CGTATGAATGGCCCCACGAGGGTGCCTGTCTCCTTTTCTAATCAATGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Phyllomedusinae	AGTATGAACGGCACCACGAAAGTTACTGTCTCCTTTTCCAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Pseudidae	AGTATCAACGGCACCACGAGGGTCACTGTCTCCTTTTCCAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Ranidae	CGTATCAACGGCACCACGAAAGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Rhinodermatidae	NGTATGAACGGCTTACGAAAGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
<i>Smilisca</i>	AGTATGAACGGCACCACGAAAGTTACTGTCTCCTTTTCTAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Sooglossidae	AGTATGAACGGCCCCACGAGGGTGCCTGTCTCCTTTTCCAATCAGTGAACTAATCTCCCCGTGAGAAAGCGGGGAT
Telmatobiinae	TGTATGAAGGCACCACGAGAAATCACACTGTCTCCTTTTCTTAGTCACTGAACTAATCTCCCCGTGAGAAAGCGGGGAT

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Discoglossidae	ATCAACATAAGACGAGAAGACCCCATGGAGCTTTAAACTCTAAG--CACCACCTTTACCTATAAACCACGGATTAAT
Leiopelmatidae	AATAACATAAGACGAGAAGACCCCATGGAGCTTAAAGACGATCTCCCCCCCCCCCCCATAGTTTAGGCAGCCA----
Pelobatidae	TTCCACCATAAGACGAGAAGACCCCATGGAGCTTTAAACTTAAAT--CACTTGCTTCAATCACTAATCCAGGGGATAAACA
Rhinophrynidae	TTCCACCATAAGACGAGAAGACCCCATGGAGCTTTAAACT--AGGACACCT-GTTTTCTTTTCAAACCAATCAGGCAT
Bufoidea	AAACCTATAAGACGAGAAGACCCCATGGAGCTTTAAACTAACAATAG-CACCTAACCTATAACAT--AAATTTCCCGAAC
Centrolenidae	AAACACATAAGACGAGAAGACCCCATGGAGCTTTAAACTACAGCAAGCATACTGTCTGACCCACTTAAATTTAGAAAA
<i>Colostethus</i>	AAATTTATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAGTAAATTTGACATCCAAATTTTATCTTCCGAGCTCC
<i>Dendrobates</i>	AAACCTATAAGACGAGAAGACCCCATGGAGCTTTAAACTAACAATAGCCTGCTATTTAACTTTATACCTTCTGAAAACT
Heleophrynidae	AAAAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTAACAATAGCCTGGCACCCTGCTTTAATAACTTTTAACTTCGGAAATTC
Hemiphractinae	ACAACCTATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAA-CAACATTTGCTAAGTACACACGAAATCTCAGGAAAT
<i>Hyla</i>	AAATCTATAAGACGAGAAGACCCCATGGAGCTTTAAACTTAAATAA-CACTTGCTTTACACAAATCTGCCCTCCAGAGCAAC
Hyperoliidae	AAGACTATAAGACGAGAAGACCCCATGGAGCTTTAAACTTATCAC-CACTTACCAAAACCCCTAAACACATCCCTGGTAAA
Leptodactylinae	AAAAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTAACAATAGCCTAACCCACTTACACAACTCCAGAGCAAT
Limnodynastinae	ATTCTTATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAACAG--CATATGCTCACCTCACCTACTCTCCGTAAGTCA
Mantellidae	TTTTTTATAAGACGAGAAGACCCCATGGAGCTTTAAACTCACCT--GCACCTGCTATTTTCTATTAACCTCTAAAC
Microhylidae	AAAAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAGTAAATTTGCTTAAATATAAAAAATTTAGAAAAATA
Myobatrachinae	GAACATATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAGTAAATTTGCTTAAATATAAAAAATTTAGAAAAATA
Pelodyadinae	AAAAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAGTAAATTTGCTTAAATATAAAAAATTTAGAAAAATA
<i>Phobobates</i>	TATCCTATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAGTAAATTTGCTTAAATATAAAAAATTTAGAAAAATA
Phyllomedusinae	AAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTTTCGGCAATTAATCTTATTTATACCCACCCAGGGGCAATCA
Pseudidae	ACAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTTTAAGCACTTTTAACTACTACATCTCTTAAATATTTCTAGC
Ranidae	AAAAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTTAAACATCAT--CAT--GCAACTCTGTCTCCATATCCCTTAAATTC
Rhinodermatidae	AACTTATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAGTAA-CAATTTGCTATAAATATCC--AAATTTAGAAAT
<i>Smilisca</i>	AAGAATATAAGACGAGAAGACCCCATGGAGCTTTAAACTAAGTAAACCTGCTTTTAAACATTTTAACTTCCGAGTACT
Sooglossidae	AAACCTATAAGACGAGAAGACCCCATGGAGCTTTCAGTAAATA-TTACATGCCCCAAAATTAACATACC-----
Telmatobiinae	GTCTCTACAAGACGAGAAGACCCCATGGAGCTTAAAGACAAAATAAACACTTGGCAGCCTACACCAACCGGTATACCCCCAC

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FIG. 1—Continued

rest of the proto-bufooid range during the split of the Indian continent from Africa (153–148 million years ago (MYA); Smith *et al.*, 1994) and remained restricted to the northern part of India + Antarctica/Australia until the disintegration of this supercontinent around 120–130 MYA (Smith *et al.*, 1994). This scenario would involve either an assumption of extinction of members of this family in India and Madagascar or a continuous

restriction of the sooglossids to the northern part of the Indian continent. Also, an older date for the separation of the Seychelles might be invoked.

Phylogenetic relationships of the Australasian Myobatrachidae to the African Heleophrynidae and of these families to the rest of the bufooids remain a phylogenetic enigma. Lynch (1973) placed the Cycloranae (now the Limnodynastinae) together with the Hele-

Discoglossidae	TTGTAATCAAGTGGACA----	TGACCCA-CTGTTTTCGGTTGGGGCGACCACGGAGAAAAACACATCCTCCGAGACGAA
Leiopelmatidae	-----	TGACTTT-CTGTCTTCGGTTGGGGCGACCACGGAGAAAAATA-ATCCTCCGAGCCGAA
Pelobatidae	CTTAACCAAGCAACGA----	TGATCT-ATGTTTTCGGTTGGGGCGACCACGGAGAAATAAATCCTCCTCGATGAA
Rhinophrynidae	AACCACTAGCTAAGCAGATCT-	GGCTCCC-TTGTTCGGTTGGGGCGACCACGGAGAAAAAAATCCTCCATGAAGAA
Bufonidae	ACTTGAATTTTACTGGTATAC-	TGACTAT-AGTTTTTGGTTGGGGTGACCAGGAGCATAATGCAACCTCCATGTTGAA
Centrolenidae	ATCTCCTTCACCTCTGCAGAA-	TGTTTGC-CAGTTTTAGGTTGGGGTGACCAGGAGAAAAAAC-ATCCTCCACATTGAA
Colostethus	CACCTCTCTGAGCATT- ----	TTATTTT- TAATTTTAGGTTGGGGCGACCACGGAGCAAAATTAACCTCCACGACGAA
Dendrobates	AAATATATCTCTAGCACTT- ----	TAATTTT- TAGTTTTAGGTTGGGGTGACCACGGAGAAAAAAATCCTCCACGTTGAA
Heleophrynidae	CATTAAAAACAAGCAGCA----	TGACCAC-AGTTTTTGGTTGGGGCGACCACGGAGAACTAAACCTGCGAGATGTA
Hemiphractinae	AATTCCTATTTTAGCCCTA----	TGATTGT-TAGTTTTAGGTTGGGGTGACCAGGAGTAAAAAACACCTCCACAATGAA
Hyla	TTACACTTACTTTAGCCTCC-	TGATTAT-TAGTTTTAGGTTGGGGTGACCAGGAGTAAAAATAACCTCCACATTGAA
Hyperoliidae	AA-----	TGACTGA-CAGTTTTCGGTTGGGGCGACCCTGGAGAAAAACAAATCCTCCAGCTGAA
Leptodactylinae	ACTTTACCTTGGC----	TTAA- TAATTTT-TAGTTTTTGGTTGGGGTGACCACGGAGAAAAAGAAACCTCCGCAATGAA
Limnodynastinae	AGCATG-----	TGACTGC-TAGTTTTCGGTTGGGGCGACCACGGAGAAACACATCCTCCGCGTTGAA
Mantellidae	TATGGAATC-----	TGCATTCTTGTTTTAGGTTGGGGTGACCAGGAGCAATAACAGCTCCACGATGAA
Microhylidae	GTGGCCC-----	TGACTGC-TAGTTTTAGGTTGGGGTGACCAGGAGTAAAAACAACTCCCAAGTAA
Myobatrachinae	AATTTATCTAAGCATT-----	TGATAAT-AGTTTTTGGTTGGGGTGACCAGGAGTAAAAAACACCTCCGCAATGAA
Pelodyridinae	-----	TGACCCA-TAGTTTTCGGTTGGGGCGACCACGGAGAAAAACCCACCTCCACGCTGAA
Phobobates	TATTTTTATTTAAGCATT-----	TTATTTT- TAGTTTTAGGTTGGGGTGACCACGGAGCAAACTAAACCTCCATGAAGAA
Phyllomedusinae	AATTTACTTCAATATTA----	TGACTGC-TAGTTTTAGGTTGGGGTGACCAGGAGTAAAAATCCTCCCAAGTAA
Pseudidae	TA-----	TGCCAA-TAGTTTTAGGTTGGGGCGACCAGGAGTAAAACTTAGCCTCCATGACAAA
Ranidae	RAGAGATG-----	TGCATGT-TAGTTTTGGTTGGGGGACCTCGGAGTATAACTTAACCTCCAAAACAAA
Rhinodermatidae	ATAAACTTATCTTTTCGCAATTTGATTAC-	TAGTTTTAGGTTGGGGTGACCAGGAGTAAAAAGTAACTCCACATTGAA
Smilisca	CAACAATAATTTAGCTTGC-----	TGATTAC-TAGTTTTAGGTTGGGGTGACCAGGAGTAAAAACAACTCCGCTTTAAA
Sooglossidae	-----	TACATAT-TCATCTTCGGTTGGGGTGACCACGGAGAAAAACAACTCCACGACAAA
Telmatobiinae	AACATCA-----	TGTTTAT-AGTCTTAGGTTGGGGCAACCGGAGTAAAAACAACTCCAGCTGAA

Discoglossidae	AGGGC-----	ATACACCCTTAACCTAGAGCCACTACTCCAA	GTAATAGAACCTCTAACCT-CCA-TGATCCAATCTT-
Leiopelmatidae	CGGGGCACACACCACCCCTTAACCAAGAGCCACAATTCCAA	GTAATAGAACCTCTAACCG-AGTCTGATCCAACTAA	
Pelobatidae	TGA-----	AACAATTTAAACCAAGAGCTACAGCTCTAA	GTATTAGAATATCTAACAT-ACATTGATCCAATCCTA
Rhinophrynidae	AGGA----	AATCTCTCTAAACCAAGAACCACTTTCTAA	GTATCAGAACTTCTGACTA-TAATTGATCCAGTCTGA
Bufonidae	AGAT-----	ACTTATCTTAGCTAAGACCTACAGCTCTAA	GCATCAATA-TATTGACAT-CCATTGACCCAATAAA
Centrolenidae	AGGGG----	CTTACCCTTAGCCAAGAGCCACACTCCAC	GCACCGAAGAAATGGACATTTAATTGACCCAAGATA
Colostethus	GGAGACTCCTT-----	CTCTTAGCTAAGAGCTACTCCTTTAA	GCATCAGCA-AACTGACCT-CTTTGACCCAATAAA
Dendrobates	CGAG-----	CCCTCCTTAGCAAAAAGTCAACAATTCAT	GCATCAACA-CATTGACCT-CCATTGACCCAATTTT
Heleophrynidae	TAGAGTAT-----	TACCCTAAGCCAAAAGCCACCGCTTAA	GCATCAACA-CCTTGACAT-ACATTGACCCAATTTA
Hemiphractinae	TAGGGCCCT-----	CCCTTAAAGCAAAAAGCCACAATTTAC	GCATCAACAAAATTGACAT-TAATTGACCCAATATT
Hyla	TGGGAA-----	AATTCCTTAGCCCTGAGCTACACTCTAA	GCACCAATA-AATTGACAT-CAATTGACCCAATATA
Hyperoliidae	CGATTA----	AAATAATCTAATGCAAGAACCAACTCAAA	CAATCAAAA-TATTGACA-AAA-TGACCCAGAATACA
Leptodactylinae	AGA-----	CTCTCCTTCTAATTTTAGGACTACTTTCTTAA	ACATCAATA-AATTGACAT-CCATTGACCCAATAAA
Limnodynastinae	AGAATTCT-----	ATTCCTAA-CTAAAAGCTACACTTTAA	GTATCAACA-CCTTGACAT-CCATTGACCCAATAAA
Mantellidae	CGGG-----	ATTACCCCTTATCTAAGAGCTACACTCTAA	GAATTAGCA-TTCTAACAT-AAATGATCCGACAAA
Microhylidae	CGA-----	CCTCCTCTAATCCGAGAGCCACCACTCAAA	GAATCAAAA-AATTGACAC-ACATTGACCCAATTTAT
Myobatrachinae	CAAGA-----	TTATTTTTTAGCTAAGAGCTCAACTCTAA	GCATCAATA-AATTGACAT-TATTGACCCAATTTA
Pelodyridinae	TGACCCCA-TCTTAGCCCAAAGCCACAGCTTCAAGCATCA	GCATC-----	CTGACAC-CCCTGACCCAATTTAA
Phobobates	ATG-----	ATATTTTTTAGCCCAAAACTACTTTTTTAA	GCATCAACA-AATTGACCT-TCATTGACCCAATATA
Phyllomedusinae	TAAATT-----	ATTTTTAGCAATAAACTACAATTTTAT	GCATCAAAA-AATTGACAC-TATTGACCCAACACT
Pseudidae	AGG-----	ACATTTCTAATCCAAGAAAGACATTTCTAA	GAATCACTA-ACCTGATGT-TTCATGACCCGACTA
Ranidae	TGGGTTA----	ACACCTTTATCCGCGAGAAACACCTCTAA	GAATTATTA-ACTAATGC-TTTTTGATCCGATAA
Rhinodermatidae	AGAA-----	ATACTTTCTAACAACAAAACACTACAATTTAT	GTATCAATA-AATTGACAT-CAATTGACCCAATTTT
Smilisca	CGGG-----	GACCCCTTAGCGATGAGCTACAATCTTAT	GCACCAATA-TATTGATTT-TCATTGACCCAATTTT
Sooglossidae	-----	CAAGCTTAAACACCAACTTAAAA	GCACCAATA-ATTTAACAC- -TTGACCCAATACAA
Telmatobiinae	CGG-----	ATACTTCTAACAACAAAAGCAACTTTTAT	GTATCATAA--ACTGACAT-AATTTGACCCAATATACT

FIG. 1—Continued

phryninae (Heleophrynidae) as a basal lineage of the Bufonoidea, suggesting that it gave rise to the Leptodactylidae which, in turn, served as a source of a vast bufonoid radiation in the Neotropics. At the same time, he proposed that the Myobatrachinae + Sooglossidae gave rise to the ranoids. Lynch's (1973) polyphyletic family, Myobatrachidae, was created to distinguish be-

tween the Old World and the New World leptodactylids. Tyler (1979) argued that no factors other than geographic distribution were used to erect the Myobatrachidae. Accordingly, he referred both the Old World and New World taxa to the Leptodactylidae, insisting that more diagnostic characters were needed for the recognition of an independent Myobatrachidae. Duell-

Discoglossidae TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCATTTTAAAGAGTCCATATCGACAAATGGGTTTACGAC
Leiopelmatidae TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCACTCCGAGAGTCCATATCGACAGCGGGTTTACGAC
Pelobatidae TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCATTTCAAGAGTCCATATCGACAAATGGGTTTACGAC
Rhinophryinae CTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCTATTTTAAAGAGTCCATATCGACAAATGGGTTTACGAC
Bufo TTGAACAACGAACCAAGTTACCTAGGGATAACAGCGCAATCCACTTCAAGAGCTCCTATCGACAAGTGGGTTTACGAC
Centrolenidae TTGATCATCGAACCAAGTTACCTAGGGATAACAGCGCAATCCATTTTAAAGAGCCCCTATCGACAAGTGGGTTTACGAC
Colostethus TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCTACTTCAAGAGCTCATATCGACAAGTGGGTTTACGAC
Dendrobates TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCTACTTCGAGAGTCTTATCGACAAGTGGGTTTACGAC
Heleophryinae TTGACCAACGAACCAAGTTACCTGGGGATAACAGCGCAAGTCAATTTCTAGAGNCCNTATCGACAGATGGGTTTACGAC
Hemiphractinae TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCCACTTCAAGAGCCCCTATCGACAAGTGGGTTTACGAC
Hyla TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCCGCTTTTAAAGAGCCCCTATCGACAAGCGGGTTTACGAC
Hyperoliidae TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCCACTTCAAGAGCCCCTATCGACAAGTGGGTTTACGAC
Leptodactylinae TTGATCAACGAACCTAGTTACCTGGGGATAACAGCGCAATCCATTTCAAGAGCCCCTATCGACAAGTGGGTTTACGAC
Limnodynastinae TCGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCACTTCAAGAGCCCCTATCGACAAGTGGGTTTACGAC
Mantellidae TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCATTTTCAAGAGCTCCTATCGACAAGTGGGTTTACGAC
Microhylidae TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCCACTTCAAGAGCTCCTATCGACAAGTGGGTTTACGAC
Myobatrachinae TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCACTTCAAGAGCTCCTATCGACAAGTGGGTTTACGAC
Pelodyadinae TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCTACTTCAAGAGCTCCTATCGACAAGTGGGTTTACGAC
Phobobates TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCTACTTCAAGAGCTCATATCGACAAGTGGGTTTACGAC
Phyllomedusinae TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCATTTCAAGAGCTCCTATCGACAAGTGGGTTTACGAC
Pseudidae TCGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCATTTTAAAGAGCTCCTATCGACAAGTGGGTTTACGAC
Ranidae TCGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCTACTTCAAGAGTTCATATCGACAAGTGGGTTTACGAC
Rhinodermatidae TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCTACTTCAAGAGCCCCTATCGACAAGTGGGTTTACGAC
Smilisca TTGATCAACGAACCAAGTTACCTGGGGATAACAGCGCAATCCGCTTCAAGAGCCCCTATCGACAAGCGGGTTTACGAC
Sooglossidae TTGATCAACGAACCAAGTTACCTAGGGATAACAGCGCAATCCATTTCAAGAGCCCCTATCGACAAGTGGGTTTACGAC
Telmatobiinae TTGATTAACGAACCAAGTTACCTAGGGATAACAGCGCAATCCACTTCAAGAGTTCATATCGAAAAGTGGGCTTACGAC

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Discoglossidae CTCGATGTTGGATCAGGGTGTCTAGTGGTGCAGCCGCCACTAAAGG-TTCGTTTGGT
Leiopelmatidae CTCGATGTTGGATCAGGACCCCTAATGGTGAAGCCGCTATTAACGTGTTTCGTTTGGT
Pelobatidae CTCGATGTTGGATCAGGGCATCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Rhinophryinae CTCGATGTTGGATCAGGGCCCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Bufo CTCGATGTTGGATCAGGGTATCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Centrolenidae CTCGATGTTGGATCAGGGTATCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Colostethus CTCGATGTTGGATCAGGGTATCCTAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Dendrobates CTCGATGTTGGATCAGGGTATCCTGGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Heleophryinae CTCGATGTTGGATCAGGGCATCCGGTGGTGNAGCCGCTACCAGCG-TTCGTTTGGT
Hemiphractinae CTCGATGTTGGATCAGGGTACCCAGTGGTGCAGCCGCTACNNNNNNNNNNNNNNNNNN
Hyla CTCGATGTTGGATCAGGGTATCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Hyperoliidae CTCGATGTTGGATCAGGGTATCCTAGCAGCGCAGCAGTTGCTCAAGG-TTCGTTNNNNN
Leptodactylinae CTCGATGTTGGATCAGGGTATCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Limnodynastinae CTCGATGTTGGATCAGGGTATCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTNNNN
Mantellidae CTCGATGTTGGATCAGGGTATCCTAGTGGTGCAGCAGCTACTAAAGNNNNNNNNNNNNNN
Microhylidae CTCGATGTTGGATCAGGGTATCCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Myobatrachinae CTCGATGTTGGATCAGGGTATCCCAGTGGTGCAGCCGCTANTAAAG-TTCGTTTGGT
Pelodyadinae CTCGATGTTGGATCAGGGTATCCAAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Phobobates CTCGATGTTGGATCAGGGTATCCTAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Phyllomedusinae CTCGATGTTGGATCAGGGTACCCAGTGGTGCAGCCGCTNNNNNNNNNNNNNNNNNN
Pseudidae CTCGATGTTGGATCAGGATACCCAGTGGTGCAGNNNNNNNNNNAAAG-TTCGTTTGGT
Ranidae CTCGATGTTGGATCAGGGTATCCTGGTGGTGCACCCGCTACTAAAG-TTCGTTTGGT
Rhinodermatidae CTCGATGTTGGATCAGGGTACCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Smilisca CTCGATGTTGGATCAGGGTACCCAGTGGTGCAGCCGCTACTAAAG-TTCGTTTGGT
Sooglossidae CTCGATGTTGGATCAGGGTATCCTAGTGGTGCACCCGCTACTAAAG-TTCGTTTGGT
Telmatobiinae CTCGATGTTGGATCAAGGTACCTGGTGGTGCAGCCGCTACCAAAG-TTCGTTTGGT

FIG. 1—Continued

man and Trueb (1986) placed a taxon composed of the Myobatrachidae + Sooglossidae and the Heleophryinae at the base of the neobatrachian radiation. Ford and Cannatella (1993) identified the Myobatrachinae and Sooglossidae as sister taxa (excluding the Limnodynastinae) on the basis of "at least five shared derived characters" of morphology. Based on our data, it is

tempting to argue that the Heleophryinae, Limnodynastinae, and Myobatrachinae (with the former two clustering) constitute a monophyletic group equal to the Myobatrachidae (*sensu* Lynch). However, given the degree of support for the appropriate nodes on the tree (Fig. 2), we can claim only that these three taxa are bufonoids ($P_c = 0.96$) but are likely to be excluded from

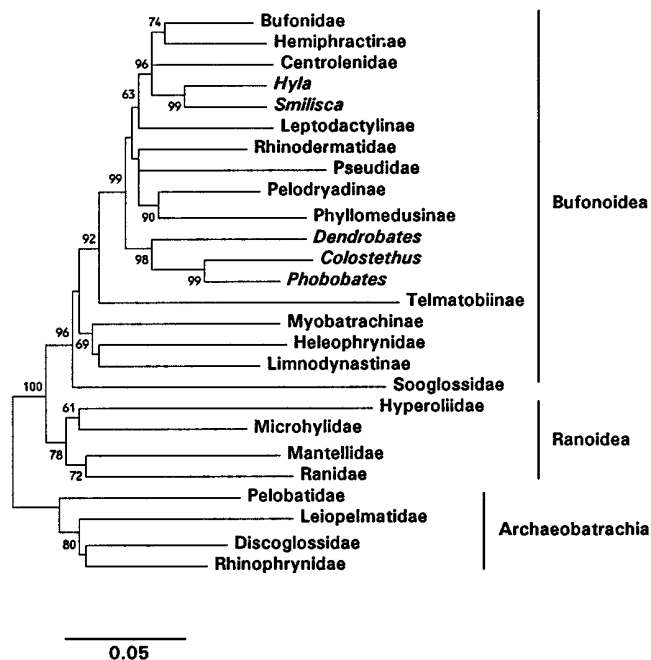


FIG. 2. Phylogenetic relationships of the Neobatrachia based on a neighbor-joining analysis of combined mitochondrial 12S and 16S rRNA gene sequences (Jukes-Cantor distances, pairwise deletion; 899 aligned sites, 465 variable). The Archaeobatrachia is used as an outgroup. The following bufonoid families were represented by more than one sample: Dendrobatidae (*Colostethus*, *Dendrobates*, and *Phobobates*), Hylidae (Hemiphractinae, Hylinae (*Hyla* and *Smilisca*), Pelodryadinae, and Phyllomedusinae), Leptodactylidae (Leptodactylinae and Telmatobiinae), and Myobatrachidae (Limnodynastinae and Myobatrachinae). Numbers on the tree represent confidence value expressed as percentages from the interior-branch test (values below 50% are not shown). The distance scale is drawn below the tree.

the neotropical radiation ($P_c = 0.92$). Their relationships with the Sooglossidae also are unclear. Based on the paleogeographic data (Smith *et al.*, 1994), speciation events giving rise to the three separate myobatrachid (*sensu* Lynch) lineages could have taken place 145–150 MYA, when Antarctica/Australia separated from Africa, thus determining the present-day distribution of these taxa. This date is in a good agreement with the one for the divergence of the Sooglossidae, considering their proximity on the tree.

Our data suggest an extensive bufonoid radiation with an apparent center in South America. Somewhat in accord with the morphology-based proposal (Lynch, 1973) we see a member of the Leptodactylidae at the base of this event. However, it is a representative of the Telmatobiinae (*Eleutherodactylus cuneatus*) which, according to Lynch's (1971) scheme, was deeply nested within the family tree and not basal to the rest of the neotropical leptodactylids as shown by our data (Fig. 2). The second member of this family which is included in the present analysis, *Lithodytes lineatus* (Leptodac-

tylinae), was placed within the neotropical bufonoids, not with the Telmatobiinae, suggesting a polyphyletic Leptodactylidae. Since the majority of the bufonoid radiation is restricted to South America and because the bufonoid diversity is the highest in this region, it is likely that initial radiation took place there after both Africa and Australia became separated. The present-day distribution of the cosmopolitan bufonids and hylines can be explained as a series of secondary dispersal events (see comments on the Pelodryadinae below).

Antarctica/Australia became disjoined from South America around 153 MYA (Smith *et al.*, 1994). They were reconnected by a narrow land bridge between 140 and 130 MYA. The area of contact between South America and Africa was more extensive; they achieved a substantial degree of separation around 125 MYA (Smith *et al.*, 1994) and became completely disjoined before 100 MYA. The several multifurcating nodes on our tree (Fig. 2) are likely a reflection of an explosive radiation taking place among the Neotropical bufonoids soon after they became restricted to South America, thus dating the event around 110–120 MYA. More extensive sampling of diverse leptodactylid lineages as well as bufonids and hylids from different localities in the Old and New Worlds will be needed to test this biogeographic hypothesis.

The Pelodryadinae is a biogeographic oddity. While the three other hylid subfamilies are predominantly South American (Hylinae is cosmopolitan), pelodryadines are restricted to Australia and New Guinea. Savage (1973) raised this subfamily to familial level, suggesting a leptodactyloid ancestry of unspecified affinity. Tyler (1979) argued for the recognition of the Australo-Papuan tree frogs as a subfamily of the Hylidae. Molecular (Maxson, 1976) and morphological (reviewed in Tyler, 1979) studies aimed at identifying a sister group of the Pelodryadinae found no apparent association between the members of this subfamily and any other bufonoid lineage tested. Our present data suggest the South American Phyllomedusinae as the most likely candidate for such association. Following the line of paleogeographic considerations above, a divergence between these two lineages had to take place immediately after or during the extensive bufonoid radiation in South America, but at or before the time of the separation of Australia. Otherwise, it would be necessary to postulate an unlikely dispersal event either across the Drake Passage or the Pacific. Considering the short internal nodes in our tree, it is possible that an ancestral phyllomedusine diverged from an ancestral pelodryadine within a few million years of the beginning of a large-scale bufonoid diversification in the Neotropics. Thus the timing of this speciation can be roughly placed at 110–120 MYA. According to the paleogeographic data (Smith *et al.*, 1994) an Antarctica/Australia supercontinent remained connected to South America from 140 until 130 MYA, after which there was an archipel-

ago link until approximately 100–105 MYA. Therefore, the inferred dates of major land mass rearrangements are within a time frame allowing all necessary speciations to occur. However, this model would require an assumption of an explosive radiation as well as rapid expansion of the ranges occupied by different lineages and, perhaps, a dispersal event through an archipelago linking South America with Antarctica/Australia. Some estimates suggest that the average temperatures in the latitudes where colonization and initial radiation are postulated to have taken place could have been about 20°C higher than at present (Barron, 1983), thus making Antarctica habitable to frogs. A split of Australia from Antarctica around 80 MYA allows ample time for the range expansion. An estimated divergence between the major groups of the Pelodyadinae (55 million years; Hutchinson and Maxson, 1987) lies well within this date.

The only other well-defined cluster ($P_c = 0.96$) within the Bufonoidea consists of the representatives of three currently recognized families: Bufonidae, Centrolenidae, and Hylidae. A significant pairing of the two hylines (*Hyla* and *Smilisca*) presents little surprise. Most workers also associate the Hylidae and Centrolenidae (Lynch, 1973; Duellman and Trueb, 1986; Ford and Cannatella, 1993). On the other hand, the suggestion of an association of the Bufonidae with the hylid subfamily Hemiphractinae is totally unexpected. We predict that wider taxonomic sampling of both the Hylidae and the Bufonidae—including additional representatives of both Old World and New World genera—will clarify relationships among these lineages. We further predict that such analyses will still show that hylid and bufonid lineages are closer to one another than they are to lineages of other bufonoid families, as already indicated in Fig. 2.

CONCLUSIONS

Our analysis of portions of mitochondrial 12S and 16S rRNA genes for additional representatives of the Neobatrachia revealed some new patterns of phylogenetic relationships within the suborder. There is additional evidence for a deep split between the Bufonoidea and Ranoidea. We are able to assign the Sooglossidae to the bufonoids; however, its sister taxon is not identified. A weakly supported group including the Australian Myobatrachidae and African Heleophryinae is placed basally to the rest of the Bufonoidea, supporting morphology-based classifications. The leptodactylid Telmatobiinae appears to lie at the base of bufonoid radiation in the Neotropics. This radiation may have served as a source for the main portion of the rich South American anuran fauna and later provided initial stocks for expansion into North America and the rest of the world. The Dendrobatidae is monophyletic and within the neotropical Bufonoidea. The Pelodyadinae and Phyllomedusinae cluster, as do the Hylinae, the Centrolenidae, the Bufonidae, and the Hemiphractinae. These arrangements suggest that the Hylidae, the Leptodactylidae, and the Myobatrachidae may not be monophyletic families. Based on the phylogenetic relationships advocated here and the paleogeographic data, we propose a scenario in which a series of vicariant events divided species ranges, resulting in speciation and accounting for the present-day species distribution. Thus, this study provides a new phylogenetic scheme for the Neobatrachia and identifies areas where additional work needs to be done. We also support our earlier assertion that extensive species sampling and an increased amount of sequence data will be required to resolve enigmatic and controversial phylogenetic relationships among the Anura.

APPENDIX

Species for which rRNA Gene Sequences Were Obtained

Family	Species	Locality	Specimen
Dendrobatidae	<i>Colostethus pratti</i>	Panama	LM 1143-A
	<i>Phobobates trivittatus</i>	Peru, Panguana	LM 739-A
Hylidae			
Hemiphractinae	<i>Gastrotheca riobambae</i>	Ecuador, San Rafael	LM 3176
Hylinae	<i>Smilisca phaeota</i>	Ecuador, Esmeraldas Province	LM 2504
Pelodyadinae	<i>Litoria cyclorhynchus</i>	Western Australia, Angerup	LM 3175
Phyllomedusinae	<i>Phyllomedusa palliata</i>	Peru, Cuzco Amazonico	LM 2010
Leptodactylidae			
Leptodactylinae	<i>Lithodytes lineatus</i>	Peru	LM 269
Myobatrachidae			
Myobatrachinae	<i>Pseudophryne guentheri</i>	Western Australia, Mt. Margaret	LM 2725

Note. LM, frozen tissue collection of LRM. Voucher specimens are available for LM 3176 (UNIMNH 94580), LM 3175 (SAMA R20141), LM 2010 (KU 205420), and LM 2725 (WAM 101218). Collections are from the University of Illinois, Museum of Natural History (UIMNH), South Australian Museum (SAMA), University of Kansas (KU), and Western Australian Museum (WAM).

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REFERENCES

- Barron, E. J. (1983). A warm, equable Cretaceous: The nature of the problem. *Earth Sci. Rev.* **19**: 305–338.
- Cabot, E. L., and Beckenbach, A. T. (1989). Simultaneous editing of multiple nucleic acid and protein sequences with ESEE. *Comput. Appl. Biosci.* **5**: 233–234.
- Cannatella, D. C., and Hillis, D. M. (1993). Amphibian relationships: Phylogenetic analysis of morphology and molecules. *Herpetol. Monogr.* **7**: 1–7.
- Dickin, A. P., Fallick, A. E., Halliday, A. N., Macintyre, R. M., and Stephens, W. E. (1986/1987). An isotopic and geochronological investigation of the younger igneous rocks of the Seychelles microcontinent. *Earth Planet. Sci. Lett.* **81**: 46–56.
- Duellman, W. E. (1975). On the classification of frogs. *Occas. Pap. Mus. Nat. Hist. Univ. Kansas* **42**: 1–14.
- Duellman, W. E. (1993). Amphibian species of the world: Additions and corrections. *Univ. Kansas Mus. Nat. Hist. Spec. Publ.* **21**: 1–372.
- Duellman, W. E., and Trueb, L. (1986). "Biology of Amphibians," McGraw-Hill, New York.
- Felsenstein, J. (1978). Cases in which parsimony or compatibility methods will be positively misleading. *Syst. Zool.* **27**: 401–410.
- Ford, L. S. (1993). The phylogenetic position of the dart-poison frogs (Dendrobatidae) among anurans: An examination of competing hypotheses and their characters. *Ethol. Ecol. Evol.* **5**: 219–231.
- Ford, L. S., and Cannatella, D. C. (1993). The major clades of frogs. *Herpetol. Monogr.* **7**: 94–117.
- Gyllensten, U. B., and Erlich, H. A. (1988). Generation of single-stranded DNA by the polymerase chain reaction and its implication to direct sequencing of the HLA-DQA locus. *Proc. Natl. Acad. Sci. USA* **85**: 7652–7656.
- Hay, J. M., Ruvinsky, I., Hedges, S. B., and Maxson, L. R. (1995). Phylogenetic relationships of amphibian families inferred from DNA sequences of mitochondrial 12S and 16S ribosomal RNA genes. *Mol. Biol. Evol.* **12**: 928–937.
- Hedges, S. B. (1994). Molecular evidence for the origin of birds. *Proc. Natl. Acad. Sci. USA* **91**: 261–264.
- Hedges, S. B., Bezy, R. L., and Maxson, L. R. (1991). Phylogenetic relationships and biogeography of xantusiid lizards, inferred from mitochondrial DNA sequences. *Mol. Biol. Evol.* **8**: 767–780.
- Hedges, S. B., Kumar, S., Tamura, K., and Stoneking, M. (1992). Human origins and analysis of mitochondrial DNA sequences. *Science* **255**: 737–739.
- Hedges, S. B., and Maxson, L. R. (1993). A molecular perspective on lissamphibian phylogeny. *Herpetol. Monogr.* **7**: 27–42.
- Hillis, D. M. (1991). The phylogeny of amphibians: Knowledge and the role of cytogenetics. In "Amphibian Cytogenetics and Systematics" (D. M. Green and S. K. Sessions, Eds.), pp. 7–31, Academic Press, San Diego.
- Hillis, D. M., Ammerman, L. K., Dixon, M. T., and de Sá, R. O. (1993). Ribosomal DNA and the phylogeny of frogs. *Herpetol. Monogr.* **7**: 118–130.
- Hutchinson, M. N., and Maxson, L. R. (1987). Phylogenetic relationships among Australian tree frogs (Anura: Hylidae: Pelodyadinae): An immunological approach. *Aust. J. Zool.* **35**: 61–74.
- Jukes, T. H., and Cantor, C. R. (1969). Evolution of protein molecules. In "Mammalian Protein Metabolism" (H. N. Munro, Ed.), pp. 21–132, Academic Press, New York.
- Kimura, M. (1980). A simple method for estimating the evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.* **16**: 111–120.
- Kumar, S., Tamura, K., and Nei, M. (1993). MEGA: Molecular Evolutionary Genetics Analysis, Version 1.01, The Pennsylvania State University, University Park, PA.
- Laurent, R. F. (1979). Esquisse d'une phylogénèse des anoures. *Bull. Soc. Zool. Fr.* **104**: 397–422.
- Lecointre, G., Philippe, H., Lê, H. L. V., and Le Guyader, H. (1993). Species sampling has a major impact on phylogenetic inference. *Mol. Phylogenet. Evol.* **2**: 205–224.
- Lynch, J. D. (1971). Evolutionary relationships, osteology, and zoogeography of leptodactyloid frogs. *Misc. Publ. Mus. Nat. Hist. Univ. Kansas* **53**: 1–238.
- Lynch, J. D. (1973). The transition from archaic to advanced frogs. In "Evolutionary Biology of the Anurans: Contemporary Research on Major Problems" (J. L. Vial, Ed.), pp. 133–182, Univ. of Missouri Press, Columbia.
- Maxson, L. R. (1976). The phylogenetic status of phyllomedusine frogs (Hylidae) as evidenced from immunological studies of their serum albumins. *Experientia* **32**: 1149–1150.
- Maxson, L. R. (1992). Tempo and pattern in anuran speciation and phylogeny: An albumin perspective. In "Herpetology: Current Research on the Biology of Amphibians and Reptiles" (K. Adler, Ed.), pp. 41–57, Society for the Study of Amphibians and Reptiles, Oxford, OH.
- Milner, A. R. (1988). The relationships and the origin of the modern amphibians. In "The Phylogeny and Classification of the Tetrapods, Vol. 1, Amphibians, Reptiles, Birds" (M. J. Benton, Ed.), pp. 59–102, Clarendon, Oxford.
- Nei, M. (1991). Relative efficiencies of different tree-making methods for molecular data. In "Phylogenetic Analysis of DNA Sequences" (M. M. Miyamoto and J. Cracraft, Eds.), pp. 90–128, Oxford Univ. Press, New York.
- Nei, M., Takezaki, N., and Sitnikova, T. (1995). Assessing molecular phylogenies. *Science* **267**: 253–255.
- Reig, O. A. (1958). Propositiones para una nueva macrosistemática de los anuros. Nota preliminar. *Physis* **21**: 109–118.
- Rzhetsky, A., and Nei, M. (1992). A simple method for estimating and testing minimum-evolution trees. *Mol. Biol. Evol.* **9**: 945–967.
- Rzhetsky, A., and Nei, M. (1994). METREE: A program package for inferring and testing minimum-evolution trees. *Comput. Appl. Biosci.* **10**: 409–412.
- Saitou, N., and Nei, M. (1987). The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **4**: 406–425.
- Sanger, F., Nicklen, S., and Coulson, A. R. (1977). DNA sequencing with chain-terminating inhibitors. *Proc. Natl. Acad. Sci. USA* **74**: 5463–5467.
- Savage, J. M. (1973). The geographic distribution of frogs: Patterns and predictions. In "Evolutionary Biology of the Anurans: Contemporary Research on Major Problems" (J. L. Vial, Ed.), pp. 351–445, Univ. of Missouri Press, Columbia.

- Sitnikova, T., Rzhetsky, A., and Nei, M. (1995). Interior-branch and bootstrap tests of phylogenetic trees. *Mol. Biol. Evol.* **12**: 319–333.
- Smith, A. G., Smith, D. G., and Funnell, B. M. (1994). "Atlas of Mesozoic and Cenozoic Coastlines," Cambridge Univ. Press, Cambridge.
- Tajima, F., and Nei, M. (1984). Estimation of evolutionary distances between nucleotide sequences. *Mol. Biol. Evol.* **1**: 269–285.
- Tamura, K., and Nei, M. (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Mol. Biol. Evol.* **10**: 512–526.
- Tyler, M. J. (1979). Herpetofaunal relationships of South America with Australia. In "The South American Herpetofauna: Its Origin, Evolution, and Dispersal" (W. E. Duellman, Ed.), pp. 73–106, University of Kansas, Lawrence.